

PHARMACOLOGY LETTERS

Accelerated Communication

INCREASED BLOOD-BRAIN BARRIER PERMEABILITY OF AMINO ACIDS IN
CHRONIC HYPERTENSION

Jian-ping Tang¹, Zhi-Qun Xu¹, Frank L. Douglas^{2,3}, Ashok Rakshit^{2,4} and Srikumaran Melethil¹

¹ Schools of Pharmacy and Medicine, University of Missouri-Kansas City, Kansas City, MO 64108

² Research Department, CIBA-GEIGY Corp., Summit, NJ 07901

(Submitted August 27, 1993; accepted September 8, 1993;
received in final form October 11, 1993)

Abstract A previous communication from this laboratory reported that brain uptake of libenzapril, a small polar molecule, was enhanced in chronic hypertension (1). The objective of this investigation was to determine if this was a more generalized phenomenon. Therefore, experiments were undertaken to examine the effect of chronic hypertension on the brain uptake of tryptophan (an amino acid with high brain permeability) and glutamic acid (one with low permeability). Brain concentrations of these two amino acids were 5- to 12-fold greater in chronic hypertensive rats, as compared to normotensive rats; the corresponding brain uptake index (BUI) values were 2- to 5-fold higher in the former group. Since blood-brain barrier transport of amino acids involve both saturable (carrier) and non-saturable (most likely, diffusion via pores) mechanisms, data from this study show that hypertension can enhance BBB transport of amino acids by affecting one or both of these pathways.

Introduction

Very little is known about the effect of chronic hypertension on the blood-brain barrier (BBB) permeability of small (M.W. < 500) polar molecules. Previous studies from our laboratory (1) showed that chronic hypertension increased brain uptake of libenzapril (LZP), a small polar molecule with a lysine side-chain. It has also been reported that BUI of tyrosine increased in chronic hypertension (2). Therefore, the objective of this study was to determine if this was a more generalized phenomenon and applied to other small molecules. As a first step in this direction, the study was extended to include two other amino acids, namely tryptophan and glutamic acid, which represent high and low permeable compound, respectively.

Materials and Methods

Radioactivity Materials: ³H₂O (1 mCi/gm), ¹⁴C-tryptophan (54.8 mCi/mmol) and ¹⁴C-glutamic acid (293.3 mCi/mmol) were purchased from New England Nuclear Corp., Boston, MA.

Corresponding Author: S. Melethil, Ph.D., M3-209, 2411 Holmes Street, Schools of Pharmacy and Medicine, University of Missouri-Kansas City, Kansas City, Missouri 64108

³ Present address: Marion Merrell Dow Inc., Kansas City, MO 64114

⁴ Present address: Hoffmann-La Roche Inc., Nurley, NJ 07110

0024-3205/93 \$6.00 + .00
Copyright © 1993 Pergamon Press Ltd All rights reserved.

Animals: Sprague-Dawley, SD (Sasco Inc., Omaha, NE) and spontaneously hypertensive, SH (Taconic Farms, Germantown, NY) rats were exposed to 12-hour light : dark cycle and housed two per cage. Rats were fed with commercial rat chow and tap water ad libitum and were allowed an acclimatization period of at least 4 days prior to experimentation.

Experimental: Male normotensive (SD) or chronic hypertensive (SH) rats weighing 280-330 gms were kept anesthetized with pentobarbital (50 mg/kg, i. p.) and maintained at 37 °C. The right femoral arteries were cannulated for blood pressure measurement. The BUI method, as reported by Oldendorf (3) with minor modifications (1), was then used to examine brain permeability (ipsilateral side) of tryptophan and glutamic acid. Briefly, the right carotid artery was isolated. A 27-G needle was bent at 90° and the tapered end was inserted into the artery, such that carotid blood flow was essentially unchanged. Following the phenylephrine infusion, the drug solution (¹⁴C-LZP, 3 mg/Kg) along with a rapidly diffusible reference compound (³H₂O) was injected via the indwelling carotid cannula. The animal was decapitated 15 sec post injection. Four brain regions, namely cerebellum, medulla and pons, cortex and a composite region consisting of the remainder of the brain were dissected and analyzed for ¹⁴C-amino acid and reference (³H₂O) compound by scintillation counting. Brain uptake index values were calculated by the relationship: $BUI = \frac{(^{14}\text{C amino acid in brain} / ^3\text{H}_2\text{O in brain})}{(^{14}\text{C-amino acid in injected solution} / ^3\text{H}_2\text{O in injected solution})}$. Tissue amino acid concentrations were corrected for residual blood (1). Regional blood volume were determined using sucrose as a non-diffusible marker (3).

Data Analysis: All values are presented as mean ± SD, n = 6 - 7. Regional BUI and brain concentrations (dpm/mg tissue) of amino acids were tested for differences by using the unpaired t test (4). Outliers were dropped using the Dixon's test for extreme values (5).

Results

Mean arterial pressures (MAP) for the two SD groups were 99 ± 12 and 100 ± 15 mmHg respectively; the corresponding MAPs for the SH groups were 183 ± 9.0 and 178 ± 30 mmHg respectively. Mean MAPs in the SH rats are significantly ($p < 0.05$) higher than those in the SD rats (Table I). Regional BUI values for tryptophan and glutamic acid are shown in Table I. As shown BUI values in SH rats were greater about 2- to 3-fold for tryptophan in all 4 regions and 3- to 5-fold for glutamic acid in all regions except the medulla and pons. In the tryptophan group, differences were significant ($p < 0.05$) for all 4 regions examined; in the glutamic acid group, only (cerebellum, cortex and composite) out of the 4 regions were significant ($p < 0.05$). Regional brain concentrations (dpm/mg tissue) of ¹⁴C-tryptophan and ¹⁴C-glutamic acid are shown in Table II. In SH rats, these values were significantly ($p < 0.05$) increased in all 4 regions except the medulla and pons; the enhancement was about 5- to 6-fold for tryptophan and 6- to 12-fold for glutamic acid excluding data for medulla and pons. Brain concentrations of glutamic acid and tryptophan in SD rats in this pooled region increased only about 20 and 60%, respectively, and these increases were not significantly different from the corresponding values in SD rats.

Discussion

A previous study from our laboratory (1) showed no differences in the BBB permeability of LZ between the two genetic strains of normotensive (SD and Wistar Kyoto) rats. Data from the present study were also comparable to values reported by Oldendorf in Wistar rats (6). Brain uptake index values for tryptophan for SD rats in this study were in good agreement with the value of 0.032 (whole brain) reported by Oldendorf; for glutamic acid, BUI values for two regions (cortex and composite) were in reasonable agreement with the reported (6) values (0.032, whole brain). Therefore, the greater brain uptake of amino acids in SH rats observed in the present study is unlikely to be due to strain differences.

Results from this study showed that brain uptake (indicated by both BUI and brain concentration) of tryptophan and glutamic acid was significantly greater in chronic hypertensive (SH) rats. Blood

brain barrier transport of tryptophan and glutamic acid involve both saturable (carrier) and non-saturable (most likely, diffusion via pores) mechanisms (6, 7). Higher BUI and brain concentration

TABLE I
Brain Uptake Index of Tryptophan and Glutamic Acid in Rats

Region	Tryptophan*		Glutamic Acid**	
	SD	SH	SD	SH
Cerebellum	0.22 ± 0.044 ^a	0.64 ± 0.15 ^a	0.15 ± 0.041 ^e	0.42 ± 0.11 ^e
Medulla + Pons	0.24 ± 0.072 ^b	0.41 ± 0.10 ^b	0.42 ± 0.11	0.52 ± 0.22
Cortex	0.29 ± 0.050 ^c	0.78 ± 0.065 ^c	0.077 ± 0.010 ^f	0.39 ± 0.13 ^f
Composite	0.27 ± 0.046 ^d	0.72 ± 0.067 ^d	0.078 ± 0.017 ^g	0.35 ± 0.089 ^g
MAP (mmHg)	99 ± 12 ^g 7	183 ± 9.0 ^g 7	100 ± 15 ^h 6	178 ± 30 ^h 7

*: 0.117 μM/kg; **: 0.0907 μM/kg
Numbers with same superscript indicate significant difference (p < 0.05)

TABLE II
Regional Brain Concentration* of ¹⁴C-Tryptophan and ¹⁴C-Glutamic Acid in Rats

Region	Tryptophan		Glutamic Acid	
	SD	SH	SD	SH
Cerebellum	21.4 ± 9.92 ^a	110 ± 98.5 ^a	17.3 ± 5.92 ^d	109 ± 45.8 ^d
Medulla + Pons	10.4 ± 1.91	16.7 ± 4.87	33.0 ± 16.5	39.7 ± 20.4
Cortex	71.8 ± 55.9 ^b	417 ± 128 ^b	25.7 ± 11.6 ^e	312 ± 160 ^e
Composite	76.0 ± 54.2 ^c	372 ± 58.0 ^c	26.3 ± 8.25 ^f	268 ± 13.0 ^f

*: dpm/mg tissue
Numbers with same superscript indicate significant difference (p < 0.05)

of these amino acids in SH rats indicated that chronic hypertension can enhance these transport mechanisms (e. g. up-regulation of amino acid transporter and/or pressure-induced opening of pores). The extent of increase for glutamic acid (about 6- to 12-fold increase) was greater than that (5- to 6-fold) for tryptophan. Since glutamic acid is more polar than tryptophan, greater brain uptake of the former in SH rats suggests that the opening of pressure-sensitive channels (or pores) may predominate over carrier up-regulation in chronic hypertension.

Increases in brain concentrations of tryptophan and glutamic acid (5- to 12-fold) in hypertensive rats were much greater than the corresponding BUI values (2- to 5-fold). This is due to the fact that

o
s
l
d
s
3
n
n
d
d
H
e

P
nt
ex
3
id
).
is

of
d-

concentration of $^3\text{H}_2\text{O}$ (the reference marker) also increased about 3- to 6-fold in SH rats (data not shown). Brain uptake index, expressed as the ratio of brain concentration of the test compound to that of the reference marker ($^3\text{H}_2\text{O}$), will increase to the same extent as brain concentrations only if the reference marker uptake remains unaltered. In this regard, it was found previously that brain uptake of LZIP was significantly increased in chronic hypertension (1), but the corresponding BUI values did not increase because uptake of LZIP and $^3\text{H}_2\text{O}$ increased to the same extent. Though LZIP has a lysine side chain, brain uptake of LZIP was unaffected in the presence of lysine (unpublished data). Therefore, it appears that increased uptake of both compounds (LZIP and $^3\text{H}_2\text{O}$) in hypertension was mediated via the same mechanism (i.e. diffusion through pressure-sensitive channels (or pores) in the endothelial cells (8)). As noted, tryptophan and glutamic acid are transported across BBB via saturable and non-saturable systems. Therefore, if hypertension enhances both pathways, then increases in brain amino acid concentrations should be higher than those of $^3\text{H}_2\text{O}$ and cause BUI values to be higher as was observed in this study.

Tryptophan and glutamic acid were chosen for this study also because they are precursors of neurotransmitters (serotonin and γ -aminobutyric acid, respectively). Since the synthesis of neurotransmitters is limited by the availability of these precursors, especially that of tryptophan, it is possible that increases in brain uptake of tryptophan and glutamic acid in chronic hypertension could influence the brain synthesis of these neurotransmitters resulting in altered brain function. Further efforts are also needed to identify the molecular basis of the altered BBB amino acid transport in chronic hypertension.

Acknowledgments

Supported in part by a research grant from CIBA-GEIGY Corporation to S. Melethil. The author would like to thank Dr. Rick Xu for helpful discussions.

References

1. J-P. TANG, A. RAKHIT, F.L. DOUGLAS and S. MELETHIL. *Pharm Res* **9** 236-243 (1992).
2. P. HATZINIKOLAOU, P. BRECHER and H. GAVRAS. *Life Sci.* **29** 1657 - 1660 (1981).
3. W.H. OLDENDORF. *Trans Am Neurol Assoc* **96** 46-50 (1971).
4. G.W. SNEDECOR and W.G. COCHRAN. *Statistical Methods*, 91-104, The Iowa State University Press, Ames, Iowa (1978).
5. S. BOLTON. *Pharmaceutical Statistics - Practical and Clinical Applications*, J. Swarbrick (Ed), 294-299, Marcel Dekker Inc., New York and Basel (1984).
6. W.H. OLDENDORF. *Am J Physiol* **221** 1629-1639 (1971).
7. W.M. PARDRIDGE. *Advances in Biochemistry*, L.J. Filer, Jr., S. Garattini, M.R. Khan, W.A. Reynolds and R.J. Wurtman (Eds), 125-137, Raven Press, New York (1979).
8. C.S. PATLAK AND O.B. PAULSON. *Microvasc. Res.* **21** 117-127 (1981).